Synthesis of Lipophilic Paramagnetic Contrast Agents

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The facile, high-yielding synthesis of a series of macrocycles 7a - k in 75–100% yield is reported. The transformation of these compounds to their carboxymethylated analogues 8a-k in 75–90% yield and subsequent gadolinium complexes 9a-k provides a series of homologous neutral paramagnetic contrast agents (PCAs) with tunable lipophilicity. Alkylated cationic intermediates **6a**–**k** are prepared in yields of 72–94% from glyoxal adduct of cyclen (5) and slight excesses of alkyl iodides. The methodology is selective for monoalkylation and amenable to large-scale synthesis.

Introduction

Since its introduction in 1978, the concept of contrast enhanced magnetic resonance imaging (CE-MRI) has evolved into a common clinical modality.^{1,2} The current generation of FDA-approved T₁ contrast agents are based predominantly on amino polyacetic acid chelates of Gd-(III) (1-3) and function by promoting the relaxivity of protons on solvating water. Although MRI was initially envisioned as a completely noninvasive technique, intravenously administered paramagnetic contrast agents (PCAs) can improve the overall quality and detail of images and allow for the unambiguous identification of pathologies that would otherwise be undetected. Numerous reports demonstrate how CE-MR images readily indicate the presence of lesions, tumors, and defects that are unobserved by conventional MRI techniques.² As a result of these successes, clinical CE-MRI applications continue to rapidly evolve. This drives the need for more efficient and pathology-specific contrast agents that increase the potential of CE-MRI as a diagnostic tool.³



The development of ligands which impart significant lipophilic character to the PCA are becoming increasingly important.⁴ Ideally, ligands that possess substantial lipophilic character should serve as more efficient carriers for the transport of the paramagnetic gadolinium ion across cell membranes. Increased hydrophobicity of the ligand should lead to plasma protein binding and some degree of hepatobiliary excretion, thereby creating the potential for liver targeting.² We envisioned that the lipophilicity could be increased by substituting one of the carboxylate arms of DOTA (2) with a hydrocarbon chain. In this paper we report the synthesis of a series of alkylated macrocycles and their Gd(III) complexes which are intended for use as PCAs. These compounds are based on the selective monoalkylation of 1,4,7,10-tetraazacyclododecane (4, cyclen) with slight excesses of simple alkyl iodides.

Original work in the area of PCA design demonstrated that the polyamino carboxylate derivatives diethylenetriaminepentaacetic acid (DTPA, 1)⁵ and 1,4,7,10-tetrakis(carboxymethyl)-1,4,7,10-tetraazacyclododecane (DO-TA)⁶ formed stable and soluble complexes with Gd(III) in water. These macrocycles have no easily accessible sites for modification; thus, subsequent studies have focused on the preparation of ligands with differentiated sidearms on the nitrogen atoms. One of the most significant results of these studies was the synthesis of 1,4,7tris(carboxymethyl)-1,4,7,10-tetraazacyclododecane (DO3A, **3**) from **4** in six steps.⁷ This ligand offered the synthetic advantage of having a free nitrogen on the macrocyclic ring available for derivatization.

Numerous examples of alkylated nitrogen-based macrocycles have appeared in the literature.⁸ Recent efforts toward alkylated PCAs have focused on derivitization of 4,⁹ which can be efficiently synthesized using the methodology of Weisman and co-workers.¹⁰ Alteration of the carbon skeleton of 4 has been investigated,¹¹ but most

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strategies have been based on N-alkylation. In addition to monoalkylation products, several instances of N1,N4dialkylated cyclen derivatives have been reported in the literature.¹² Handel and co-workers used highly reactive alkyl halides and a silicon-based protection scheme to synthesize the monomethylated and monobenzylated derivatives of **4**.¹³ Benzyl, propyl, and nonyl cyclen have been prepared in 56, 63, and 48% yield, respectively, via their molybdenum tricarbonyl complexes.¹⁴ Other reports have demonstrated that the use of a 5-10 mol excess of 4 over the alkyl halide yields monoalkylation products.¹⁵ Recently, Kimura and co-workers synthesized hexadecylcyclen from dioxotetramine and 1-bromohexadecane through a multiple-step, low-yielding sequence.¹⁶

In addition to alkylated cyclen derivatives, reports of alkylated DO3A analogues have also appeared in the literature. Tweedle and co-workers reported the cyclenbased synthesis of methyl DO3A,⁷ and Chang has determined the stability constants of propyl DO3A with several alkaline-earth metals.¹⁷ A report on EPR studies of gadolinium pentyl-DO3A (Gd-DOTA-P) in phospholipid bilayers has also recently appeared.¹⁸

Results and Discussion

Cyclen (4) was synthesized through methods reported earlier by Weisman and co-workers. It has been previously shown that reaction of cyclen with glyoxal yields bridged macrocycle 5.19 Subsequent alkylation of 5 with alkyl iodides over the course of 1-4 days in toluene resulted in precipitation of the monoalkylated iodo salts 6a-k. In general, the use of longer-chain alkyl iodides required longer reaction times. We observed 1,7-dialkylation products in the case of the two most reactive halides (MeI and BnBr), even when less than 1 mol equiv of halide was employed. These results are consistent with those in the literature.²⁰

Salts **6f**–**k** were found to be slightly soluble in toluene due to their significant lipophilic character, resulting in

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Figure 1.

partial precipitation of oily amorphous residues which were subsequently purified through recrystallization. We attribute the driving force of the alkylation reaction to the precipitation of the monoalkylated cationic products from solution. Addition of a large excess of halide did not significantly hasten the course of the reaction; thus, a 1.5:1 ratio of alkyl iodide to 5 was employed in each case. The tetradecyl salt **6i** was formed in significantly higher yield than its counterparts. The origin of this effect is unknown. All iodides were pure before use; furthermore, distillation of alkyl iodides from copper mesh immediately prior to use did not significantly change the yields of subsequent alkylations.

Salts 6a-k exhibited extremely complex ¹H NMR spectra resulting from symmetry breaking of the glyoxal ring adduct. Spectra obtained at 500 MHz lacked the necessary dispersion required for full assignment of ring protons, but ¹H NMR spectra could be completely assigned at 750 MHz. HMBC, HMQC, and COSY data were used to elucidate the structure of 6c. The presence of pseudoaxial and pseudoequatorial protons on each of the four rings leads to the observation of numerous complex multiplet signals in the region from 2.40 to 4.40 ppm. Assignment of the individual ethylene fragments was achieved through the use of COSY and HMQC data, followed by positioning in the fused ring structure on the basis of long-range correlations from the HMBC experiment. In the specific case of the butyl salt **6c**, secondorder signals at 1.50, 1.60, and 1.80 ppm were observed from prodiastereotopic protons on the butyl side chain. Higher chain salts were solved by analogy due to the similarity of spectra throughout the series.

Pentyl salt 6d was recrystallized from acetone and petroleum ether over 24 h to yield crystals suitable for X-ray diffraction. This compound crystallizes in the space group *P*1 with two molecules per unit cell (Figure 1). This structure clearly shows the hydrogen atoms of the bridging ethylene unit protruding in a cisoid fashion from the bowl-shaped fused ring system. Alkylation of glyoxal adduct may occur at N1 or N2, leading to formation of a pair of enantiomers related to each other in the unit cell by an inversion center. We also observe the iodine counterion and a single solvent molecule of acetone for each molecule of 6d in the unit cell.

Formation of alkylated cyclen derivatives (7a-k) was achieved in good yields through hydrolysis of salts 6a-k

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in 20% (w/v) aqueous KOH solution at 60 °C. The course of these reactions was monitored by ¹H NMR, showing full conversion in most cases after 4 days. The resulting water-insoluble oily residues were extracted into chloroform, yielding the hydrolysis products 7a-k. Extraction of cyclen adducts 7e-k revealed the presence of a small amount of unknown impurity, which could not be removed by chromatographic methods or recrystallization. Dissolution of the residues in ethanol, followed by treatment with concentrated HCl, resulted in the formation of the respective hydrochloride salts of 7e-k. Recrystallization of these salts from ethanol, followed by neutralization with NaOH and extraction into chloroform, resulted in removal of the unknown impurities.

Hydrolysis of salts 7j and 7k resulted in the initial formation of white sebacious spheres in the reaction mixture after 24 h. These structures persisted for 3 days, after which the familiar oily residue separated from the reaction mixtures. This white substance was found to be insoluble in aqueous and organic solvents and thus was not identifiable by standard NMR techniques. The mechanism of this reaction is not currently known. We postulate a hydrolysis mechanism that proceeds via nucleophilic attack on one of the bridgehead carbons of the salt **6**, resulting in the formation of a nitrogen ylide 10 (Scheme 2). Subsequent loss of formaldehyde and formate anion affords the monosubstituted cyclen derivative 7. Attempted hydrolysis of propargylated salt 61 resulted in elimination of the propargyl arm and regeneration of glyoxal adduct 5. This demonstrates that 5 is stable under basic conditions and suggests that a positively charged nitrogen atom is a prerequisite for the hydrolysis. Generation of the ylide 10 is slow compared to deprotonation of the terminal alkyne hydrogen and subsequent elimination of the propargyl side chain. On the basis of this observation, we propose the formation of the nitrogen ylide 10 as the product-determining step in the hydrolysis. Reaction of 7a - k with chloroacetic acid and NaOH in aqueous solution yielded DO3A analogues 8a-k. During purification, 8a-h were found to be watersoluble, whereas compounds 8g-k showed limited solubility in water.

Additional solubility tests showed these compounds to be only moderately soluble in organic solvents due to formation of aggregates. Attempted characterization of 8g-k by standard NMR techniques produced spectra with low signal-to-noise due to the poor solubility of these compounds in most deuterated solvents. These compounds are currently being tested to determine their



Table 1. Synthesis of Alkylated DO3A Analogues (% yield)

entry	R	6	7	8	9
а	C_2H_5	92	92	88	100
b	C_3H_7	91	95	81	100
с	C_4H_9	84	100	87	100
d	C ₅ H ₁₁	76	100	78	100
e	C ₆ H ₁₃	79	84	83	100
f	C ₈ H ₁₇	77	79	90	100
g	$C_{10}H_{21}$	72	75	86	100
ň	$C_{12}H_{25}$	85	82	89	100
i	$C_{14}H_{29}$	94	85	80	100
i	C ₁₆ H ₃₃	74	84	77	100
ĸ	C ₁₈ H ₃₇	77	78	75	100
1	C_2H_2	94	0	-	_

critical micellular concentrations (CMC); these results will be reported separately.

Preparation of the Gd(III) complexes of 8a-k was achieved using Gd₂O₃ in aqueous solution as reported in the literature.⁷ After removal of excess Gd(OH)₃ via filtration, each solution was concentrated in vacuo to give 9a-k as glassy solids. These compounds showed solubility traits similar to their gadolinium-free counterparts and exhibited decreased solubility in both organic and aqueous solvent when the lipophilic side chain contained more than eight carbons.

Conclusion

We have prepared a series of new lipophilic Gd(III) complexes for use as paramagnetic contrast agents. This synthesis of compounds with varying lengths of alkyl side chains has resulted in the creation of potential PCAs with a tunable lipophilicity. Our methodology allows for selective monoalkylation of cyclen using slight excesses of alkyl halide in excellent yield. This procedure is restricted to the use of sidearms that do not easily undergo base-catalyzed processes. Complexes **9a**–**k** are under investigation as candidates for use as paramagnetic contrast agents for magnetic resonance imaging.

Experimental Section

General Methods. Et₂O (Mallinckrodt) was freshly distilled from benzophenone ketyl prior to use. 1-Iodotetradecane was obtained from Chemica Alta LTD, Edmonton, Alberta, Canada. All other alkyl iodides were obtained from Aldrich Chemical Co. Cyclen (4)¹⁰ and glyoxal adduct of cyclen (5)¹⁹ were prepared according to known procedures. Dowex 50W-8X cation-exchange resin was obtained from Bio-Rad Laboratories, Richmond, CA. Other reagents were purchased from commercial sources and used as obtained. Elemental analyses were performed by the University of Illinois Microanalysis Laboratory.

General Procedure for the Reaction of 5 with Alkyl Halides. A solution of *cis*-13–1,4,7,10-Tetraazatetracyclo- $[5.5.2.0^{.4.14}0^{10.13}]$ tetradecane 5 in toluene (20 mL) and alkyl iodide (1.5 equiv) was stirred at 60 °C for 4 d under N₂. In the case of salts **6a**–**e**, the reaction mixture was cooled to room temperature and filtered to yield white precipitates which required no further purification. In the case of salts **6f**–**k**, the reaction mixture was concentrated in vacuo, giving a yellow solid which was recrystallized from ether/chloroform to give pure **6f–k**.

(1*RS*,13*SR*,14*RS*)-1-Ethyl-4,7,10-triaza-1-azoniatetracyclo[5.5.2.0.^{4,14}0^{10,13}]tetradecane Iodide (6a). Yield (92%). ¹H NMR (500 MHz, CDCl₃) 1.52 (t, J = 7.0 Hz, 3H) 2.43 (ddd, J = 10.2, 7.9, 6.3 Hz, 1H), 2.67 (m, 1H), 2.70 (m, 1H) 2.80 (ddd, J = 12.5, 8.0, 6.1 Hz, 1H) 2.85 (td, J = 12.2, 3.2 Hz, 1H) 2.92 (ddd, J = 13.2, 11.7, 2.3 Hz, 1H) 3.04 (ddd, J = 11.1, 3.0, 1.9 Hz, 1H) 3.20 (m, 3H) 3.27 (td, J = 8.0, 4.5 Hz, 1H) 3.45 (td, J = 8.2, 2.5 Hz, 1H) 3.49 (d, J = 2.9, 1H) 3.78 (m, 3H) 3.91 (td, J = 13.9, 7.2 Hz, 1H) 4.19 (td, J = 10.5, 6.3 Hz, 1H) 4.20 (J = 2.8 Hz, 1H) 4.43 (ddd, J = 11.6, 10.1, 3.0 Hz, 1H). ¹³C NMR (500 MHz, CDCl₃) 9.4 43.5 47.9 48.0 48.4 49.1 52.0 53.0 57.3 61.4 72.8 83.5. HRFAB calcd for C₁₂H₂₃N₄I: C, 41.15 (40.94); H, 6.62 (6.70); N, 16.00 (15.67).

(1*RS*,13*SR*,14*RS*)-1-Propyl-4,7,10-triaza-1-azoniatetracyclo[5.5.2.0^{4,14}.0^{10,13}]tetradecane Iodide (6b). Yield (91%). ¹H NMR (500 MHz, CDCl₃) 1.13 (t, J = 7.2 Hz, 3H) 1.92 (m, 2H) 2.44 (ddd, J = 9.3, 8.0, 5.9 Hz, 1H) 2.66 (m, 1H) 2.69 (m, 1H) 2.79 (ddd, J = 12.3, 8.0, 6.1 Hz, 1H) 2.83 (td, J = 11.1, 2.6 Hz, 1H) 2.93 (ddd, J = 13.6, 11.4, 2.5 Hz, 1H) 3.03 (dt, J = 12.1, 2.0 Hz, 1H) 3.19 (m, 3H) 3.26 (td, J = 8.0, 4.6 Hz, 1H) 3.44 (td, J = 8.2, 2.5 Hz, 1H) 3.51 (d, J = 2.8, 1H) 3.74 (td, J = 12.5, 5.2, 1H) 3.80 (m, 3H) 4.05 (td, J = 12.1, 5.1 Hz, 1H) 4.16 (d, J = 2.5 Hz, 1H) 4.51 (ddd, J = 12.4, 10.0, 2.8 Hz, 1H) ¹³C NMR (500 MHz, CDCl₃) 10.9 17.3 43.6 47.8 48.1 48.3 49.1 52.0 57.7 58.8 62.0 72.1 83.6. HRFAB calcd for C1₃H₂₅N₄I: C, 42.86 (42.95); H, 6.92 (6.90); N, 15.38 (14.99).

(1*RS*,13*SR*,14*RS*)-1-Butyl-4,7,10-triaza-1-azoniatetracyclo[5.5.2.0.^{4,14}0^{10,13}]tetradecane Iodide (6c). Yield (84%). ¹H NMR (750 MHz, CDCl₃) 0.99 (t, J = 7.2 Hz, 3H), 1.49 (m, 1H), 1.57 (m, 1H), 1.82 (m, 1H), 2.42 (ddd, J = 9.7, 8.1, 6.0 Hz, 1H), 2.66 (m, 1H), 2.68 (m, 1H) 2.78 (ddd, J = 12.5, 8.0, 5.9 Hz, 1H) 2.82 (td, J = 11.7, 2.8 Hz, 1H) 2.90 (ddd, J = 13.6, 10.5, 3.0 Hz, 1H) 3.01 (ddd, J = 13.6, 3.1, 1.9 Hz, 1H) 3.19 (m, 3H) 3.24 (td, J = 8.1, 4.6 Hz, 1H) 3.43 (td, J = 8.2, 2.6 Hz, 1H) 3.49 (d, J = 2.9, 1H) 3.73 (td, J = 12.4, 5.3 Hz, 1H) 3.78 (m, 3H) 4.03 (td, J = 12.2, 4.8 Hz, 1H) 4.19 (d, J = 2.8 Hz, 1H) 4.41 (ddd, J = 11.9, 10.1, 3.0 Hz, 1H). ¹³C NMR (750 MHz, CDCl₃) 13.7 19.7 25.5 43.6 47.8 48.0 48.3 49.0 51.9 57.3 57.7 61.9 72.167 83.5. HRFAB calcd for C1₄H₂₇N₄I: C, 44.44 (44.52); H, 7.20 (7.37); N, 14.81 (14.59).

(1*RS*,13*SR*,14*RS*)-1-Pentyl-4,7,10-triaza-1-azoniatetracyclo[5.5.2.0^{4,14}.0^{10,13}]tetradecane Iodide (6d). Yield (76%). ¹H NMR (500 MHz, CDCl₃) 0.90 (t, J = 7.1 Hz, 3H) 1.40 (m, 3H) 1.52 (m, 1H), 1.84 (m, 1H) 2.43 (ddd, J = 10.0, 8.1, 6.3 Hz, 1H), 2.66 (m, 1H), 2.70 (m, 1H) 2.80 (ddd, J = 12.2, 8.1, 6.1 Hz, 1H) 2.82 (td, J = 12.6, 3.7 Hz, 1H) 2.88 (ddd, J = 13.6, 10.3, 4.9 Hz, 1H) 3.02 (ddd, J = 11.3, 2.7, 1.6 Hz, 1H) 3.20 (m, 3H) 3.26 (td, J = 8.1, 4.6 Hz, 1H) 3.44 (td, J = 8.3, 2.8 Hz, 1H) 3.49 (d, J = 2.9, 1H) 3.74 (ddd, J = 13.0, 10.9, 6.6 Hz, 1H) 3.79 (m, 3H) 4.04 (td, J = 10.7, 6.2 Hz, 1H) 4.22 (d, J = 2.8 Hz, 1H) 4.44 (ddd, J = 11.8, 10.0, 2.9 Hz, 1H). ¹³C NMR (500 MHz, CDCl₃) 13.8 22.3 23.4 28.3 43.7 47.9 48.1 48.3 49.1 51.0 57.5 57.8 61.9 72.2 83.5. HRFAB calcd for $C_{15}H_{29}N_4$ 265.2392, found 265.2392. Anal. Calcd (found) for $C_{15}H_{29}N_4$ I: C, 45.92 (46.20); H, 7.45 (7.47); N, 14.28 (13.94).

(1*RS*,13*SR*,14*RS*)-1-Hexyl-4,7,10-triaza-1-azoniatetracyclo[5.5.2.0^{4,14}.0^{10,13}]tetradecane iodide (6e). Yield (79%). ¹H NMR (500 MHz, CDCl₃) 0.87 (t, J = 7.0 Hz, 3H) 1.32 (m, 4H) 1.43 (m, 1H) 1.54 (m, 1H), 1.83 (m, 1H) 2.41 (ddd, J =9.8, 7.8, 5.9 Hz, 1H), 2.66 (m, 1H), 2.70 (m, 1H) 2.80 (ddd, J =12.3, 8.0, 5.9 Hz, 1H) 2.83 (td, J = 11.1, 3.4 Hz, 1H) 2.87 (ddd, J = 13.7, 9.8, 4.0 Hz, 1H) 3.02 (ddd, J = 11.1, 3.3, 1.9 Hz, 1H) 3.19 (m, 3H) 3.26 (td, J = 8.1, 4.6 Hz, 1H) 3.44 (td, J = 8.2, 2.8 Hz, 1H) 3.47 (d, J = 2.9, 1H) 3.73 (ddd, J = 12.8, 10.5, 8.2 Hz, 1H) 3.79 (m, 3H) 4.05 (td, J = 12.8, 7.9 Hz, 1H) 4.20 (d, J =2.8 Hz, 1H) 4.44 (ddd, J = 12.6, 9.9, 2.9 Hz, 1H) 4.20 (MR (500 MHz, CDCl₃) 13.9 22.3 23.6 25.9 31.2 43.6 47.8 48.0 48.3 49.0 51.9 57.4 57.8 61.9 72.2 83.4. HRFAB calcd for C₁₆H₃₁N₄I: C, 47.29 (47.31); H, 7.69 (7.65); N, 13.79 (13.77).

(1RS,13SR,14RS)-1-Octyl-4,7,10-triaza-1-azoniatetracyclo[5.5.2.04,14.010,13]tetradecane Iodide (6f). Yield (77%). ¹H NMR (500 MHz, CDCl₃) 0.84 (t, J = 7.1 Hz, 3H) 1.30 (m, 8H) 1.43 (m, 1H) 1.51 (m, 1H), 1.82 (m, 2H) 2.45 (ddd, J = 10.4, 8.3, 4.4 Hz, 1H), 2.68 (m, 1H), 2.71 (m, 1H) 2.80 (ddd, J = 13.9, 8.0, 3.4 Hz, 1H) 2.86 (td, J = 11.1, 3.2 Hz, 1H) 2.91 (ddd, J = 11.5, 8.7, 4.0 Hz, 1H) 3.03 (ddd, J = 10.8, 3.3, 2.0 Hz, 1H) 3.21 (m, 3H) 3.27 (td, J = 8.0, 4.8 Hz, 1H) 3.45 (td, J= 8.4, 2.9 Hz, 1H) 3.55 (d, J = 2.8, 1H) 3.74 (ddd, J = 13.0,10.0, 7.0 Hz, 1H) 3.79 (m, 3H) 4.02 (td, J = 12.7, 7.9 Hz, 1H) 4.23 (d, J = 2.8 Hz, 1H) 4.40 (ddd, J = 12.0, 9.9, 3.0 Hz, 1H). ¹³C NMR (500 MHz, CDCl₃) 14.0 22.5 23.6 26.3 28.9 29.2 31.6 43.6 47.8 48.1 48.3 49.1 51.9 57.4 57.8 57.8 61.9 72.2 83.4. HRFAB calcd for $C_{18}H_{35}N_4$ 307.2862, found 307.2862. Anal. Calcd (found) for C₁₈H₃₅N₄I: C, 49.77 (49.99); H, 8.12 (8.23); N, 12.90 (12.93).

(1RS,13SR,14RS)-1-Decyl-4,7,10-triaza-1-azoniatetracyclo[5.5.2.0.4,14010,13]tetradecane Iodide (6g). Yield (72%). ¹H NMR (500 MHz, CDCl₃) 0.86 (t, J = 7.0 Hz, 3H) 1.24 (m, 10H) 1.35 (m, 2H) 1.49 (m, 2H), 1.82 (m, 2H) 2.43 (ddd, J= 9.7, 8.0, 5.6 Hz, 1H), 2.68 (m, 1H), 2.71 (m, 1H) 2.82 (ddd, J =12.4, 8.1, 5.9 Hz, 1H) 2.86 (td, J = 11.7, 2.8 Hz, 1H) 2.88 (ddd, J = 13.9, 8.4, 5.9 Hz, 1H) 3.04 (ddd, J = 11.1, 4.76, 1.9 Hz, 1H) 3.20 (m, 3H) 3.27 (td, J = 8.2, 4.6 Hz, 1H) 3.45 (td, J =8.2, 2.8 Hz, 1H) 3.49 (d, J = 2.7, 1H) 3.75 (ddd, J = 12.7, 10.0, 7.0 Hz, 1H) 3.79 (m, 3H) 4.05 (td, J = 12.9, 5.8 Hz, 1H) 4.22 (d, J = 2.9 Hz, 1H) 4.44 (ddd, J = 12.9, 10.1, 3.0 Hz, 1H). ¹³C NMR (500 MHz, CDCl₃) 14.0 22.5 23.6 26.3 29.1 29.2 29.3 29.3 31.7 43.6 47.8 48.0 48.2 49.0 51.9 57.5 57.7 61.9 72.1 83.3. HRFABMS calcd for C₂₀H₃₉N₄ 335.3175, found 335.3176. Anal. Calcd (found) for C₂₀H₃₉N₄I: C, 51.94 (51.75); H, 8.50 (8.34); N, 12.11 (12.10).

(1RS,13SR,14RS)-1-Dodecyl-4,7,10-triaza-1-azoniatetracvclo[5.5.2.0.4,14010,13]tetradecane Iodide (6h). Yield (85%). ¹H NMR (500 MHz, CDCl₃) 0.87 (t, J = 7.1 Hz, 3H) 1.25 (m, 14H) 1.36 (m, 2H) 1.50 (m, 2H), 1.81 (m, 2H) 2.41 (ddd, J= 9.6, 7.7, 5.4 Hz, 1H), 2.69 (m, 1H), 2.72 (m, 1H) 2.82 (ddd, J= 12.0, 8.2, 6.1 Hz, 1H) 2.83 (td, J = 11.7, 2.9 Hz, 1H) 2.85 (ddd, J = 12.3, 8.6, 5.3 Hz, 1H) 3.04 (ddd, J = 11.3, 4.5, 1.6 Hz, 1H) 3.21 (m, 3H) 3.28 (td, J = 8.1, 4.7 Hz, 1H) 3.44 (td, J = 9.0,3.2 Hz, 1H) 3.45 (d, J = 2.8, 1H) 3.76 (td, J = 12.9, 5.6 Hz, 1H) 3.80 (m, 3H) 4.10 (td, J = 12.0, 5.1 Hz, 1H) 4.22 (d, J = 12.02.8 Hz, 1H) 4.44 (ddd, J = 12.3, 9.9, 3.2 Hz, 1H). ¹³C NMR (500 MHz, CDCl₃) 14.1 17.3 22.6 23.7 26.3 29.3 29.3 29.5 29.5 31.8 43.7 47.9 48.2 48.4 49.1 52.0 57.4 58.0 62.0 72.3 83.3. HRFABMS calcd for $C_{22}H_{43}N_4$ 363.3488, found 363.3486. Anal. Calcd (found) for C₂₂H₄₃N₄I: C, 53.86 (53.86); H, 8.83 (8.73); N, 11.42 (11.17).

(1*RS*,13*SR*,14*RS*)-1-Tetradecyl-4,7,10-triaza-1-azoniatetracyclo[5.5.2.0.^{4,14}0^{10,13}]tetradecane Iodide (6i). Yield (94%). ¹H NMR (500 MHz, CDCl₃) 0.87 (t, J = 7.1 Hz, 3H) 1.24 (m, 18H) 1.35 (m, 2H) 1.49 (m, 2H), 1.81 (m, 2H) 2.40 (ddd, J = 9.4, 7.9, 5.6 Hz, 1H), 2.69 (m, 1H), 2.71 (m, 1H) 2.81 (ddd, J = 12.3, 7.9, 5.9 Hz, 1H) 2.83 (td, J = 10.7, 2.9 Hz, 1H) 2.85 (ddd, J = 12.8, 9.6, 5.9 Hz, 1H) 3.04 (ddd, J = 11.4, 5.1, 1.9 Hz, 1H) 3.20 (m, 3H) 3.28 (td, J = 8.2, 4.7 Hz, 1H) 3.44 (d, J = 2.9 Hz, 1H) 3.45 (td, J = 8.0, 2.5 1H) 3.75 (td, J = 12.7, 4.9 Hz, 1H) 3.80 (m, 3H) 4.08 (td, J = 12.6, 4.9 Hz, 1H) 4.22 (d, J = 2.7 Hz, 1H) 4.44 (ddd, J = 13.2, 10.0, 3.1 Hz, 1H). ¹³C NMR (500 MHz, CDCl₃) 14.1 22.6 23.7 26.4 28.5 29.3 29.3 29.4 29.5 29.6 29.6 31.9 43.7 47.9 48.1 48.3 49.1 52.0 57.5 62.0 72.2 83.3 HRFAB calcd for C₂₄H₄₇N₄ 391.3801, found 391.3803. Anal. Calcd (found) for C₂₄H₄₇N₄I: C, 55.59 (55.34); H, 9.14 (9.31); N, 10.80 (10.55).

(1RS.13SR.14RS)-1-Hexadecvl-4.7.10-triaza-1-azoniatetracyclo[5.5.2.0.4,14010,13]tetradecane Iodide (6j). Yield (74%). ¹H NMR (500 MHz, CDCl₃) 0.86 (t, J = 7.1 Hz, 3H) 1.25 (m, 22H) 1.35 (m, 2H) 1.48 (m, 2H), 1.81 (m, 2H) 2.41 (ddd, J = 9.5, 7.9, 5.7 Hz, 1H), 2.68 (m, 1H), 2.71 (m, 1H) 2.81 (ddd, J = 12.9, 9.6, 6.6 Hz, 1H) 2.83 (m, 1H) 2.86 (ddd, J =12.8, 9.6, 6.6 Hz, 1H) 3.03 (ddd, J = 11.0, 4.5, 2.0 Hz, 1H) 3.19 (m, 3H) 3.27 (td, J = 8.0, 4.5 Hz, 1H) 3.42 (td, J = 8.1, 2.6 Hz, 1H) 3.46 (d, J = 2.8 1H) 3.75 (td, J = 11.4, 6.0 Hz, 1H) 3.80 (m, 3H) 4.08 (td, J = 12.5, 5.0 Hz, 1H) 4.22 (d, J = 2.6Hz, 1H) 4.44 (ddd, J = 12.1, 10.0, 3.3 Hz, 1H). ¹³C NMR (500 MHz, CDCl₃) 14.0 22.6 23.7 26.3 29.3 29.5 29.5 29.6 29.6 31.8 43.743 47.9 48.1 48.4 49.1 52.0 57.4 57.9 62.0 72.3 83.3. HRFAB calcd for C₂₆H₅₁N₄ 419.4114, found 419.4121. Anal. Calcd (found) for C₂₆H₅₁N₄I: C, 57.13 (57.04); H, 9.40 (9.43); N, 10.24 (9.93).

(1RS,13SR,14RS)-1-Octadecyl-4,7,10-triaza-1-azoniatetracyclo[5.5.2.0.4,14010,13]tetradecane Iodide (6k). Yield (77%). ¹H NMR (500 MHz, CDCl₃) 0.86 (t, J = 7.1 Hz, 3H) 1.24 (m, 26H) 1.35 (m, 2H) 1.48 (m, 2H), 1.81 (m, 2H) 2.42 (ddd, J = 9.6, 7.2, 5.7 Hz, 1H), 2.68 (m, 1H), 2.71 (m, 1H) 2.81 (ddd, J = 12.0, 7.9, 5.9 Hz, 1H) 2.83 (m, 1H) 2.87 (ddd, J =3.4, 9.5, 6.7 Hz, 1H) 3.03 (td, J = 11.3, 2.3 Hz, 1H) 3.19 (m, 3H) 3.26 (td, J = 8.0, 4.6 Hz, 1H) 3.45 (td, J = 8.2, 2.6 Hz, 1H) 3.47 (d, J = 2.7 1H) 3.74 (td, J = 11.2, 5.6 Hz, 1H) 3.79 (m, 3H) 4.08 (td, J = 11.1, 5.1 Hz, 1H) 4.22 (d, J = 2.8 Hz, 1H) 4.44 (ddd, J = 11.8, 10.2, 2.8 Hz, 1H). ¹³C NMR (500 MHz, CDCl₃) 14.1 22.6 23.7 26.3 28.5 29.3 29.4 29.5 29.6 29.6 29.7 30.5 31.9 33.5 43.7 48.0 48.2 48.4 49.1 52.0 57.4 58.0 62.0 72.4 83.3. HRFAB calcd for C₂₈H₅₅N₄ 447.4427, found 447.4425. Anal. Calcd (found) for C₂₈H₅₅N₄I: C, 58.52 (58.17); H, 9.65 (9.71); N, 9.75 (9.45).

(1*RS*,13*SR*,14*RS*)-1-Propargyl-4,7,10-triaza-1-azoniatetracyclo[5.5.2.0^{4,14}.0^{10,13}]tetradecane Iodide (6l). Yield (94%). ¹H NMR (500 MHz, D₂O, benzene as internal reference) 2.30 (ddd, $J = 10.0 \, 8.6, 6.2 \, Hz, 1H$), 2.40 (td, $J = 11.7, 3.0 \, Hz,$ 1H), 2.64 (td, $J = 11.7, 3.6 \, Hz, 1H$), 2.71 (m, 3H), 2.77 (ddd, $J = 12.1, 7.7, 6.1 \, Hz, 1H$), 3.10 (m, 4H), 3.15 (td, J = 8.5, 4.3Hz, 1H), 3.38 (td, $J = 8.1, 2.5 \, Hz, 1H$), 3.41 (d, $J = 2.7 \, Hz,$ 1H), 3.61 (td, $J = 12.8, 3.5 \, Hz, 1H$), 3.74 (dt, $J = 12.5, 8.4 \, Hz,$ 1H), 3.86 (d, J = 13.1, 1H), 3.94 (d, $J = 2.4 \, Hz, 1H$), 3.98 (dd, $J = 12.9, 9.9, 2.9 \, Hz, 1H$), 4.39 (AB, Jab = 16.5, $\Delta \nu = 40.4,$ 2H). ¹³C NMR (500 MHz, D₂O, benzene as internal reference) 43.7, 47.5, 47.9, 48.0, 48.1, 48.9, 51.1, 59.1, 63.1, 70.4, 71.4, 81.7 82.0. HRFAB calcd for C₁₃H₂₁N₄ 233.1766, found 233.1765. Anal. Calcd (found) for C₁₃H₂₁N₄I: C, 43.27 (43.40); H, 5.86 (5.77); N, 15.53 (15.77).

General Procedure for the Hydrolysis of Alkylated Salts. An aqueous solution of 20% (w/v) KOH (25 mL) and alkylated salt **6** was stirred at 60 °C under N₂ for 4 d. The solution was cooled to room temperature and extracted with CHCl₃ (3 × 30 mL). The organic layers were combined, dried (Na₂SO₄), and concentrated in vacuo to yield the alkylated cyclen derivative as a yellow oil. Compounds **7a**–**d** required no further purification. Compounds **7e**–**k** were purified as follows. To a solution of the oil in ethanol (3 mL) was added concentrated HCl (10 mL) to yield the tetrahydrochloride salt as a white precipitate. The solid was recrystallized from ethanol, dissolved in 20% (w/v) KOH (20 mL), and extracted with CHCl₃ (3 × 30 mL). The organic layers were combined, dried (Na₂SO₄), and concentrated to yield the alkylated cyclen derivative.

1-Ethyl-4,7,10-tetraazacyclododecane (7a). Yield (92%). ¹H NMR (500 MHz, CDCl₃) 1.03 (t, J = 7.1 Hz, 3H), 2.52 (t, J = 7.1 Hz, 2H), 2.54 (t, J = 5.1, 4H), 2.59 (t, J = 4.9 Hz, 4H), 2.64 (t, J = 4.8 Hz, 4H), 2.79 (t, J = 5.0 Hz, 4H). ¹³C NMR (500 MHz, CDCl₃) 11.9 45.2 45.9 47.0 48.0 50.7. HRCIMS calcd for C₁₀H₂₄N₄ 201.2079, found 201.2079 (M + H). Anal. Calcd (found) for C₁₀H₂₈N₄Cl₄: C, 34.70 (34.77); H, 8.15 (8.28); N, 16.19 (16.00).

1-Propyl-1,4,7,10-tetraazacyclododecane (7b). Yield (95%). ¹H NMR (500 MHz, CDCl₃) 0.80 (t, J = 7.3 Hz, 3H), 1.39 (sext, J = 7.3 Hz, 2H), 2.28 (t, J = 7.2, 2H), 2.42 (t, J = 4.9 Hz, 4H), 2.47 (t, J = 5.2 Hz, 4H), 2.53 (t, J = 5.1 Hz, 4H), 2.68 (t, J = 5.1 Hz, 4H). ¹³C NMR (500 MHz, CDCl₃) 11.7 20.2 44.9 45.9 46.8 41.3 56.1. HRCIMS calcd for C₁₁H₂₆N₄ 215.2236, found 215.2232 (M + H). Anal. Calcd (found) for C₁₁H₃₀N₄Cl₄: C, 36.67 (37.01); H, 8.40 (8.61); N, 15.56 (15.59).

1-Butyl-1,4,7,10-tetraazacyclododecane (7c). Yield (100%). ¹H NMR (500 MHz, CDCl₃) 0.88 (t, J = 7.3 Hz, 3H), 1.29 (sext, J = 7.3 Hz, 2H), 1.43 (quin, J = 7.2, 2H), 2.38 (t, J = 7.2 Hz, 2H), 2.49 (t, J = 4.9 Hz, 4H), 2.54 (t, J = 5.1 Hz, 4H), 2.60 (t, J = 5.1 Hz, 4H) 2.75 (t, J = 5.0, 4H). ¹³C NMR (500 MHz, CDCl₃) 13.9 22.2 29.4 45.2 45.9 47.0 51.4 54.3. HRCIMS calcd for $C_{12}H_{28}N_4$ 229.2331 found 229.2330 (M + H). Anal. Calcd (found) for $C_{12}H_{32}N_4Cl_4$: C, 38.51 (38.69); H, 8.61 (9.00); N, 14.97 (15.11).

1-Pentyl-1,4,7,10-tetraazacyclododecane (7d). Yield (100%). ¹H NMR (500 MHz, CDCl₃) 0.87 (t, J = 7.1 Hz, 3H), 1.28 (m., 4H), 1.45 (quin, J = 7.2, 2H), 2.41 (t, J = 7.2 Hz, 2H), 2.54 (t, J = 4.9 Hz, 4H), 2.59 (t, J = 5.1 Hz, 4H), 2.65 (t, J = 5.2 Hz, 4H) 2.79 (t, J = 5.1, 4H). ¹³C NMR (500 MHz, CDCl₃) 14.0 22.4 26.7 29.5 45.2 45.8 47.0 51.3 54.5. HRCIMS calcd for C₁₃H₃₀N₄ 243.2438, found 243.2437 (M + H). Anal. Calcd (found) for C₁₃H₃₄N₄Cl₄: C, 40.22 (40.60); H, 8.83 (8.94); N, 14.43 (14.71).

1-Hexyl-1,4,7,10-tetraazacyclododecane (7e). Yield (84%). ¹H NMR (500 MHz, CDCl₃) 0.81 (t, J = 7.0 Hz, 3H), 1.22 (m., 6H), 1.41 (quin, J = 7.1, 2H), 2.34 (t, J = 7.2 Hz, 2H), 2.46 (t, J = 5.0 Hz, 4H), 2.51 (t, J = 5.0 Hz, 4H), 2.57 (t, J = 5.0 Hz, 4H) 2.72 (t, J = 5.0, 4H). ¹³C NMR (500 MHz, CDCl₃) 14.0 15.6 22.4 26.8 29.5 45.6 45.9 46.9 51.3 54.3. HRCIMS calcd for C₁₄H₃₂N₄ 257.2732., found 257.2733 (M + H). Anal. Calcd (found) for C₁₄H₃₆N₄Cl₄: C, 41.80 (41.67); H, 9.02 (8.88); N, 13.93 (14.33).

1-Octyl-1,4,7,10-tetraazacyclododecane (7f). Yield (79%). ¹H NMR (500 MHz, CDCl₃) 0.85 (t, J = 7.2 Hz, 3H), 1.25 (m., 10H), 1.44 (quin, J = 7.4, 2H), 2.38 (t, J = 7.2 Hz, 2H), 2.50 (t, J = 5.3 Hz, 4H), 2.54 (t, J = 5.2 Hz, 4H), 2.60 (t, J = 5.1 Hz, 4H) 2.76 (t, J = 5.2, 4H). ¹³C NMR (500 MHz, CDCl₃) 14.0 22.6 27.2 27.4 29.3 29.4 31.8 44.9 45.9 46.8 51.4 54.4 HRCIMS calcd for C₁₆H₃₆N₄ 285.3022, found 285.3033 (M + H). Anal. Calcd (found) for C₁₆H₄₀N₄Cl₄: C, 44.66 (45.00); H, 9.36 (9.71); N, 13.02 (13.00).

1-Decyl-1,4,7,10-tetraazacyclododecane (7g). Yield (75%). ¹H NMR (500 MHz, CDCl₃) 0.85 (t, J = 7.2 Hz, 3H), 1.25 (m, 10H), 1.44 (quin, J = 7.4, 2H), 2.38 (t, J = 7.2 Hz, 2H), 2.50 (t, J = 5.3 Hz, 4H), 2.54 (t, J = 5.2 Hz, 4H), 2.60 (t, J = 5.1 Hz, 4H) 2.76 (t, J = 5.2, 4H). ¹³C NMR (500 MHz, CDCl₃) 14.1 22.6 27.2 27.4 29.3 29.5 29.6 31.8 32.5 45.2 46.0 47.1 51.5 54.5. HRCIMS calcd for C₁₈H₄₀N₄ 313.3331, found 313.3327 (M + H). Anal. Calcd (found) for C₁₈H₄₄N₄Cl₄: C, 47.16 (47.41); H, 9.67 (9.87); N, 12.23 (11.90).

1-Dodecyl-1,4,7,10-tetraazacyclododecane (7h). Yield (82%).¹H NMR (500 MHz, CDCl₃) 0.86 (t, J = 7.1 Hz, 3H), 1.24 (m., 18H), 1.45 (quin, J = 7.2, 2H), 2.39 (t, J = 7.3 Hz, 2H), 2.52 (t, J = 5.5 Hz, 4H), 2.58 (t, J = 5.1 Hz, 4H), 2.64 (t, J = 5.2 Hz, 4H) 2.79 (t, J = 5.1, 4H). ¹³C NMR (500 MHz, CDCl₃) 14.0 22.6 27.2 29.3 29.5 29.5 29.6 29.6 31.8 45.3 46.0 47.1 52.0 52.1. HRCIMS calcd for C₂₀H₄₄N₄ 341.3644, found 341.3640 (M + H).

1-Tetradecyl-1,4,7,10-tetraazacyclododecane (7i). Yield (85%). ¹H NMR (500 MHz, CDCl₃) 0.87 (t, J = 7.2 Hz, 3H), 1.25 (m., 22H), 1.44 (quin, J = 7.1, 2H), 2.37 (t, J = 7.4 Hz, 2H), 2.55 (t, J = 5.2 Hz, 4H), 2.58 (t, J = 5.3 Hz, 4H), 2.61 (t, J = 5.2 Hz, 4H) 2.77 (t, J = 5.1, 4H). ¹³C NMR (500 MHz, CDCl₃) 14.1 22.6 27.2 27.5 27.6 29.1 29.3 29.4 29.5 29.6 29.6 29.7 31.9 45.2 46.0 47.1 51.5 52.1. HRCIMS calcd for C₂₂H₄₈N₄ 369.3957, found 369.3951 (M + H).

1-Hexadecyl-1,4,7,10-tetraazacyclododecane (7j). Yield (84%). ¹H NMR (500 MHz, CDCl₃) 0.87 (t, J = 7.1 Hz, 3H), 1.24 (m., 22H), 1.45 (quin, J = 7.2, 2H), 2.38 (t, J = 7.3 Hz, 2H), 2.51 (t, J = 5.3 Hz, 4H), 2.56 (t, J = 5.1 Hz, 4H), 2.62 (t, J = 5.0 Hz, 4H) 2.76 (t, J = 5.0, 4H). ¹³C NMR (500 MHz, CDCl₃) 14.0 22.6 27.2 27.4 27.4 27.6 29.3 29.5 29.6 29.6 31.8 45.2 46.0 47.0 51.481 54.550. HRCIMS calcd for C₂₄H₅₂N₄ 397.4271, found 397.4262 (M + H).

1-Octadecyl-1,4,7,10-tetraazacyclododecane (7k). Yield (78%). ¹H NMR (500 MHz, CDCl₃) 0.86 (t, J = 7.0 Hz, 3H), 1.24 (m, 30H), 1.44 (quin, J = 6.9, 2H), 2.38 (t, J = 7.1 Hz, 2H), 2.51 (t, J = 5.1 Hz, 4H), 2.56 (t, J = 5.0 Hz, 4H), 2.62 (t, J = 5.0 Hz, 4H) 2.77 (t, J = 5.0, 4H) ¹³C NMR (500 MHz, CDCl₃) 14.1 22.6 27.3 27.5 27.8 29.3 29.4 29.5 29.6 29.7 29.7 29.8 31.9 45.1 46.1 47.0 51.5 54.5. HRCIMS calcd for C₂₆H₅₆N₄ 425.4471, found 425.4479 (M + H).

General Procedure for Carboxymethylation. A solution of monoalkylated cyclen **7** in water (10 mL), chloroacetic acid (3.5 equiv), and NaOH (3.5 equiv) was stirred under N₂ at 80 °C for 48 h, maintaining the pH between 9 and 10 by the addition of 1 M NaOH as needed. The solution was cooled to room temperature and concentrated in vacuo to give a yellow solid which was acidified to pH 2.5 with 1 M HCl and loaded onto Dowex 50W-X8 cation-exchange resin in the H⁺ form. The column was eluted with 1 L water to remove salts and excess chloroacetic acid, followed by elution with 1 L of 0.5 M NH₃ to collect the desired product. The NH₃ eluent was concentrated in vacuo and triturated with ethanol to give the respective triacetic acid **8** as a yellow solid. The solid was recrystallized as the tetrahydrochloride salt from acetonitrile and ethanol for characterization purposes.

1-Ethyl-1,4,7,10-tetraazacyclododecane-1,4,7-triacetic Acid (8a). Yield (88%). ¹H NMR (500 MHz, D₂O, benzene as internal reference) δ 1.57 (3H, t, J = 7.1, H_a), 2.94 (10H, br s, H_b), 3.05 (4H, m, H_c), 3.20 (4H, s, H_d), 3.28 (4H, m, H_e) 3.58 (2H, s, H_f). ¹³C NMR (125 MHz, D₂O, benzene as internal reference) δ 16.0 48.5 48.7 48.9 50.1 51.1 53.5 55.2 56.4 170.2 178.1. HRFABMS calcd for C₁₆H₃₁N₄O₆ 375.2244, found 375.2243 (M + H). Anal. Calcd (found) for C₁₆H₃₄N₄O₆-Cl₄: C, 36.94 (36.80); H, 6.59 (6.41); N, 10.77 (10.71).

1-Propyl-1,4,7,10-tetraazacyclododecane-1,4,7-triacetic Acid (8b). Yield. (81%). ¹H NMR (500 MHz, D₂O, benzene as internal reference) δ 1.10 (3H, t, J= 7.3, H_a), 1.55 (2H, sext, J= 7.2 Hz, H_b), 3.00 (10H, br s, H_c), 3.04 (4H, m, H_d), 3.19 (4H, s, H_e), 3.27 (4H, m, H_d) 3.59 (2H, s, H_g). ¹³C NMR (125 MHz, D₂O, benzene as internal reference) δ 13.1 21.3 47.6 48.3 48.6 50.1 51.3 54.1 55.5 57.1 171.1 176.7. HRFABMS calcd for C₁₇H₃₃N₄O₆ 389.2400, found 389.2402 (M + H). Anal. Calcd (found) for C₁₇H₃₆N₄O₆Cl₄: C, 38.21 (37.99); H, 6.79 (6.64); N, 10.49 (10.85).

1-Butyl-1,4,7,10-tetraazacyclododecane-1,4,7-triacetic Acid (8c). Yield (87%). ¹H NMR (500 MHz, D₂O, benzene as internal reference) δ 0.77 (3H, t, J = 7.3 Hz, H_a), 1.20 (2H, sext, J = 7.4, H_b), 1.52 (2H, quin, J = 7.2 Hz, H_c), 2.97 (10H, br s, H_e), 3.03 (4H, m, H_f), 3.18 (4H, s, H_g), 3.26 (4H, m, H_h), 3.56 (2H, s, H_i). ¹³C NMR (125 MHz, D₂O, benzene as internal reference) δ 12.6 19.1 24.2 48.6 48.8 48.9 50.0 51.1 53.4 55.3 56.3 170.1 177.6. HRFABMS calcd for C₁₈H₃₅N₄O₆ 403.2556 (M + H). Anal. Calcd (found) for C₁₈H₃₈N₄O₆Cl₄: C, 39.43 (39.11); H, 6.98 (6.64); N, 10.22 (10.16).

1-Pentyl-1,4,7,10-tetraazacyclododecane-1,4,7-triacetic Acid (8d). Yield (78%). ¹H NMR (500 MHz, D₂O, benzene as internal reference) δ 0.85 (3H, t, J = 7.2 Hz, H_a), 1.20 (4H, m, H_b), 1.66 (2H, quin, J = 7.3 Hz, H_c) 3.05 (10H, br s, H_e), 3.18 (4H, m, H_f), 3.20 (4H, s, H_g), 3.33 (4H, m, H_h) 3.60 (2H, s, H_i). ¹³C NMR (125 MHz, D₂O, benzene as internal reference) δ 12.8 21.3 21.8 27.8 48.7 48.9 49.9 51.1 53.6 55.3 56.3 170.1 177.4. HRFAB calcd for C₁₉H₃₇N₄O₆ 417.2713, found 417.2713 (M + H). Anal. Calcd (found) for C₁₉H₄₀N₄O₆Cl₄: C, 40.58 (40.19); H, 7.17 (7.37); N, 9.96 (10.02).

1-Hexyl-1,4,7,10-tetraazacyclododecane-1,4,7-triacetic Acid (8e). Yield (83%). ¹H NMR (500 MHz, D₂O, benzene as internal reference) δ 0.86 (3H, t, J = 7.2 Hz, H_a), 1.20 (6H, m, H_b), 1.66 (2H, quin, J = 7.3 Hz, H_c) 3.10 (10H, br s, H_e), 3.25 (4H, m, H_f), 3.34 (4H, s, H_g), 3.43 (4H, m, H_h) 3.70 (2H, s, H_i). 13 C NMR (125 MHz, D₂O, benzene as internal reference) δ 13.1 21.6 21.8 22.0 25.4 26.4 30.4 30.9 48.8 51.2 51.3 52.5 53.7 55.3 56.6. HRFABMS calcd for $C_{20}H_{39}N_4O_6$ 431.2869, found 431.2869 (M + H). Anal. Calcd (found) for $C_{20}H_{42}N_4O_6Cl_4$: C, 41.68 (41.77); H, 7.34 (6.99); N, 9.72 (10.01).

1-Octyl-1,4,7,10-tetraazacyclododecane-1,4,7-triacetic Acid (8f). Yield (90%). ¹³C NMR (125 MHz, D₂O, benzene as internal reference) δ 13.2 21.7 22.0 25.3 25.6 28.0 28.1 28.1 30.8 48.7 48.7 48.9 49.9 51.1 53.6 55.2 55.3 56.4. HRFABMS calcd for C₂₂H₄₃N₄O₆ 458.3214, found 459.3217 (M + H). Anal. Calcd (found) for C₂₂H₄₆N₄O₆Cl₄: C, 43.71 (44.01); H, 7.67 (7.81); N, 9.27 (8.90).

 $\begin{array}{l} 1\mbox{-} Decyl\mbox{-} 1,4,7,10\mbox{-} tetraazacyclododecane\mbox{-} 1,4,7\mbox{-} triacetic Acid (8g).\mbox{21 Yield (86\%). HRFABMS calcd for $C_{24}H_{47}N_4O_6$ 487.3496, found 487.3495 (M + H). Anal. Calcd (found) for $C_{24}H_{50}N_4O_6Cl_4$: C, 45.58 (45.64); H, 7.96 (7.87); N, 8.86 (8.46). \end{array}$

1-Dodecyl-1,4,7,10-tetraazacyclododecane-1,4,7-triacetic Acid (8h). Yield (89%). HRFABMS calcd for $C_{26}H_{51}N_4O_6$ 515.3698, found 515.3699 (M + H).

1-Tetradecyl-1,4,7,10-tetraazacyclododecane-1,4,7-triacetic Acid (8i). Yield (80%). FABMS calcd for $C_{28}H_{55}N_4O_6$ 543.4023, found 543.4023 (M + H).

1-Hexadecyl-1,4,7,10-tetraazacyclododecane-1,4,7-triacetic Acid (8j). Yield (77%). HRFABMS calcd for $C_{30}H_{59}N_4O_6$ 571.4232, found 571.4232 (M + H).

1-Octadecyl-1,4,7,10-tetraazacyclododecane-1,4,7-triacetic Acid (8k). Yield (75%). HRFABMS calcd for $C_{32}H_{63}N_4O_6$ 599.4531, found 599.4532 (M + H).

General Procedure for the Formation of Gadolinium Complexes. A solution of the ligand **8** in water (2 mL) was brought to pH 3 with glacial acetic acid. Gd₂O₃ (1.1 equiv based on Gd) was added and the solution stirred under N₂ for 24 h at 80 °C. The cloudy solution was cooled to room temperature, filtered through a 0.2- μ m filter, and concentrated in vacuo to yield **9a**-**k** as glassy solids, which were subsequently recrystallized from acetonitrile and water.

[1-Ethyl-4,7,10-tris(carboxymethyl)tetraazacyclododecanato]gadolinium (9a). Yield (100%). HRFABMS calcd for $C_{16}H_{28}N_4O_6$ Gd 530.0905, found 530.0907 (M + H). Anal. Calcd (found) for $C_{16}H_{27}N_4O_6$ Gd· H_2O ·2.5NaOAc: C, 33.57 (33.53); H, 4.89 (5.14); N, 7.45 (7.43).

[1-Propyl-4,7,10-tris(carboxymethyl)tetraazacyclododecanato]gadolinium (9b). Yield (100%). HRFABMS calcd for $C_{17}H_{30}N_4O_6$ Gd 544.1176, found 544.1176 (M + H). Anal. Calcd (found) for $C_{17}H_{29}N_4O_6$ Gd·3.5H₂O·1.5 NaOAc: C, 32.96 (32.81); H, 5.60 (5.41); N, 7.68 (7.74).

[1-Butyl-4,7,10-tris(carboxymethyl)tetraazacyclododecanato]gadolinium (9c). Yield (100%). HRFABMS calcd for $C_{18}H_{32}N_4O_6Gd$ 558.1449, found 558.1450 (M + H). Anal. Calcd (found) for $C_{18}H_{31}N_4O_6Gd$ ·2.5H₂O·1.5 NaOAc: C, 34.80 (34.96); H, 5.63 (5.69); N, 7.73 (7.73).

[1-Pentyl-4,7,10-tris(carboxymethyl)tetraazacyclododecanato]gadolinium (9d). Yield (100%). HRFABMS calcd for $C_{19}H_{34}N_4O_6Gd$ 572.1719, found 592.1718 (M + H). Anal. Calcd (found) for $C_{19}H_{33}N_4O_6Gd\cdot 4H_2O\cdot 1.5NaOAc:$ C, 33.71 (33.66); H, 5.85 (5.75); N, 7.14 (7.49).

⁽²¹⁾ Insolubility of compounds with alkyl chains longer than eight carbons rendered elemental analysis impossible.

Anal. Calcd (found) for $C_{24}H_{43}N_4O_6Gd$ ·6NaOAc: C, 38.16 (38.54); H, 5.42 (5.73); N, 4.94 (4.95).

[1-Tetradecyl-4,7,10-tris(carboxymethyl)tetraazacyclododecanato]gadolinium (9i). Yield (100%). HRFABMS calcd for $C_{28}H_{52}N_4O_6Gd$ 698.4146, found 698.4140 (M + H).

[1-Hexadecyl-4,7,10-tris(carboxymethyl)tetraazacyclododecanato]gadolinium (9j). Yield (100%). HRFABMS calcd for $C_{30}H_{56}N_4O_6Gd$ 726.4687, found 726.4689 (M + H).

[1-Octadecyl-4,7,10-tris(carboxymethyl)tetraazacyclododecanato]gadolinium (9k).Yield (100%). HRFABMS calcd for $C_{32}H_{60}N_4O_6Gd$ 754.5221, found 754.5222 (M + H).

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Supporting Information Available: X-ray structure tables for **6d**; ¹H NMR and ¹³C NMR spectra for **6a–l**, **7a–k**, **8a–e**; ¹³C NMR data for **8f**; spectral assignments for **6a–k**, **7a–k**. This material is available free of charge via the Internet at http://pubs.acs.org.

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